

DOI: <https://doi.org/10.23868/gc562791>

Вклад 25-гидроксихолестерина в перекрёстное взаимодействие иммунной и нервной систем

Г.Ф. Закирьянова^{1,2}, А.Н. Ценцевичский¹, А.Р. Гиниатуллин^{1,2}, С.М.Ф. Нгомси³,
Е.А. Кузнецова¹, А.М. Петров^{1,2}

¹ Казанский институт биохимии и биофизики — обособленное структурное подразделение Федерального исследовательского центра «Казанский научный центр Российской академии наук», Казань, Российская Федерация;

² Казанский государственный медицинский университет, Казань, Российская Федерация;

³ Казанский федеральный университет, Казань, Российская Федерация

АННОТАЦИЯ

25-гидроксихолестерин (25ГХ) образуется из холестерина при участии фермента холестерин-25-гидроксилазы, экспрессия которой, как и уровень 25ГХ, значительно увеличивается в макрофагах, дендритных клетках и микроглии при воспалительной реакции. В свою очередь 25ГХ действует на многие иммунные клетки, модулируя течение воспалительной реакции и препятствуя проникновению вирусов в клетки. Накапливаются данные об участии 25ГХ в регуляции синаптической передачи как в центральной, так и периферической нервной системах. Учитывая повышенную продукцию 25ГХ не только при воспалении, но при ряде нейродегенеративных заболеваний (болезни Альцгеймера и боковом амиотрофическом склерозе), этот гидроксихолестерин может иметь значение в адаптации синаптической активности к воспалительным условиям, а также участвовать в патогенезе нейродегенеративных заболеваний и формировании синаптических дисфункций. Мишенями 25ГХ в нервной системе являются глутаматные NMDA-рецепторы, печёночные X-рецепторы и эстрогеновые рецепторы. К тому же 25ГХ может напрямую влиять на свойства синаптических мембран, изменяя формирование мембранных микродоменов (липидных рафтов) — компартментов, где сосредоточены белки, важные в синаптической пластичности. Текущие данные указывают на то, что эффекты 25ГХ сильно зависят от концентрации и «контекста» (норма, патология, наличие воспалительной реакции), в котором исследуется его действие.

В данном мини-обзоре мы сфокусировались на ключевых аспектах действия 25ГХ как локального регулятора гомеостаза холестерина и как паракриной молекулы, реализующей влияние воспаления на процессы нейротрансмиссии в центральной и периферической нервной системе.

Ключевые слова: синаптическая передача; 25-гидроксихолестерин; скелетная мышца; везикула; липидные рафты; нейромедиатор.

Как цитировать:

Закирьянова Г.Ф., Ценцевичский А.Н., Гиниатуллин А.Р., Нгомси С.М.Ф., Кузнецова Е.А., Петров А.М. Вклад 25-гидроксихолестерина в перекрёстное взаимодействие иммунной и нервной систем // Гены и клетки. 2023. Т. 18, № 4. С. 269–280. DOI: <https://doi.org/10.23868/gc562791>

DOI: <https://doi.org/10.23868/gc562791>

Contribution of 25-hydroxycholesterol to the cross-interaction of the immune and nervous systems

Guzalia F. Zakyrganova^{1, 2}, Andrei N. Tsentsevitsky¹, Arthur R. Giniatullin^{1, 2},
Sonia M.F. Nghomsi³, Eva A. Kuznetsova¹, Alexey M. Petrov^{1, 2}

¹ Kazan Institute of Biochemistry and Biophysics, Federal Research Center “Kazan Scientific Center of RAS”, Russian Academy of Sciences, Kazan, Russian Federation;

² Kazan State Medial University, Kazan, Russian Federation;

³ Kazan Federal University, Kazan, Russian Federation

ABSTRACT

25-hydroxycholesterol (25HC) is produced from cholesterol by cholesterol-25-hydroxylase, and its expression, similar to the 25HC level, increases significantly in macrophages, dendritic cells, and microglia during an inflammatory reaction. In turn, 25HC acts on many immune cells; therefore, it can modulate the course of the inflammatory reaction and prevent the penetration of viruses into cells. Data are accumulating about the involvement of 25HC in the regulation of synaptic transmission in both the central and peripheral nervous systems. 25HC production is increased not only during inflammation but in certain neurodegenerative diseases, such as Alzheimer’s disease and amyotrophic lateral sclerosis; thus, this hydroxycholesterol can be important in the adaptation of synaptic activity to inflammatory conditions, pathogenesis of neurodegenerative diseases, and formation of synaptic dysfunctions. The targets of 25HC in the nervous system are glutamate NMDA receptors, liver X-receptors, and estrogen receptors. 25HC can also directly influence the properties of synaptic membranes by changing the formation of membrane microdomains (lipid rafts) where proteins, which are important for synaptic plasticity, are clustered. Current data indicate that the effects of 25HC strongly depend on its concentration and “context” (norm, pathology, and presence of an inflammatory reaction) in which the effect of 25HC is being investigated. This minireview focused on the key aspects of the action of 25HC as both a local regulator of cholesterol homeostasis and a paracrine molecule that realizes the influence of inflammation on neurotransmission processes in the central and peripheral nervous systems.

Keywords: synaptic transmission; 25-hydroxycholesterol; skeletal muscle; vesicle; lipid rafts; neurotransmitter.

To cite this article:

Zakyrganova GF, Tsentsevitsky AN, Giniatullin AR, Nghomsi SMF, Kuznetsova EA, Petrov AM. Contribution of 25-hydroxycholesterol to the cross-interaction of the immune and nervous systems. *Genes & cells*. 2023;18(4):269–280. DOI: <https://doi.org/10.23868/gc562791>

INTRODUCTION

Cholesterol is found in large quantities in the plasma membranes of cells (approximately 20–30 mole % [1]), where it maintains the rigidity, fluidity, and permeability of the bilipid layer. Cholesterol in combination with glycosphingolipids forms nanometer dynamic lipid microdomains that serve as a scaffold for various proteins. In addition, cholesterol has a great affinity for many transmembrane proteins (ion channels and receptors) and can modulate their activity, thereby affecting intracellular processes. In addition to its role in maintaining the structure and functioning of membranes, cholesterol is transformed into various oxysterols by enzymatic and oxidative reactions. Oxysterols have structural and functional diversity. Recent studies have indicated that oxysterols have target proteins in both the central and peripheral nervous systems. Particularly, transcription factors (liver X receptors, retinoid orphan receptors, estrogen receptor- α , glucocorticoid receptors, and sterol regulatory element-binding protein [SREBP 1]), receptors associated with G-proteins (GPR183, GPR17, CXCR2, SMO, and SLO1), ion channels (P2X7 and NMDA receptors), cell adhesion molecules ($\alpha_5\beta_1$ -integrin), and oxysterol-binding proteins (ORP8) are stimulated or modulated by oxysterols [2–9].

Cytochrome P450 46A1 (CYP46A1) is one of the main enzymes of the brain that transform cholesterol into an oxidized form. This enzyme catalyzes the formation of 24-hydroxycholesterol (24HC) [10], which passes through the blood–brain barrier and enters the bloodstream with subsequent delivery to the liver [11, 12]. This pathway is responsible for removing 75–85% and 40–50% of excess cholesterol from the human and mouse brains, respectively [13, 14].

In addition to 24HC, 25-hydroxycholesterol (25HC) participates in controlling the level of brain cholesterol. Unlike 24HC, 25HC is produced in the brain in significantly smaller quantities, is not brain-specific, and is produced in the periphery. Its flow from the brain to the periphery and from the periphery to the brain is determined by the concentration gradient. 25HC is produced by the enzyme cholesterol-25-hydroxylase (CH25H). The *CH25H* gene is interferon-induced, and interferon γ and interferon type I can rapidly and STAT1-dependently enhance the expression of CH25H [15]. In general, CH25H expression increases rapidly in inflammation in macrophages, dendritic cells, and microglia. 25HC activates LX receptors, which can lead to the formation of interferon- γ [16], increasing CH25H expression. During inflammation, the blood plasma concentrations of 25HC can reach 200 ng/mL [17], whereas under normal conditions, its content ranges from 2 to 30 ng/mL [18].

25-HYDROXYCHOLESTEROL AS A REGULATOR OF CHOLESTEROL HOMEOSTASIS

25-hydroxycholesterol is an affecting regulator of cholesterol biosynthesis. By binding to LX receptors, 25HC increases the expression of cholesterol transporters ABCA1 and ABCG1 [19], which provide the outflow of excess cholesterol and phospholipids from cells [20]. An additional mechanism of action of 25HC in the regulation of cholesterol homeostasis is the inhibition of cholesterol synthesis through the binding of 25HC to insulin-induced gene 1 (INSIG). INSIG is a protein of the endoplasmic reticulum, which undergoes conformational changes by binding to 25HC and further interacts with SREBP cleavage-activating protein (SCAP), eventually forming the INSIG–SCAP–SREBP-2 complex. As part of this complex, SREBP-2 remains in an inactive form that prevents the formation of the active transcription factor SREBP-2, which enhances the expression of cholesterol biosynthesis genes [21, 22]. The results of the analysis of the interaction of INSIG and 25HC revealed the ubiquitination and degradation of 3-hydroxy-3-methylglutaryl-coenzyme A reductase, which catalyzes the key stage of cholesterol synthesis [23]. Thus, the more cholesterol the cells contain, the more intensively it is synthesized into 25HC, which inhibits the synthesis of cholesterol and promotes its removal from the cell; as a result, the cholesterol level in the cell normalizes.

25-HYDROXYCHOLESTEROL AND IMMUNE RESPONSE

Numerous data have revealed the mechanisms of action of 25HC in immune cells in the presence of inflammation. 25HC has a multidirectional effect on modulating the immune response by inhibiting or increasing the production of inflammatory cytokines. 25HC can induce the secretion of proinflammatory cytokines and chemokines, for example, interleukin (IL)-1 β , IL-6, IL-8, CC motif chemokine ligand 5 and macrophage colony-stimulating factor. On the contrary, 25HC suppresses inflammation by reducing the activity of inflammasomes [24, 25]. When Toll-like receptor 4 (TLR4) of macrophages is activated, there is an increase in CH25H expression and 25HC synthesis, which leads to the suppression of IL-2 production that is necessary for B-cell proliferation [17]. An increase in macrophage survival and a decrease in the innate immune response were found to be dependent on Toll receptors during the stimulation of LX receptors of macrophages by 25HC, which led to a decrease in the production of IL-6, IL-1 β , monocyte chemoattractant

proteins (MCP-1 and MCP-3), inducible nitric oxide synthase, and matrix metalloproteinase-9 [26]. As mentioned above, interferon stimulates CH25H expression. In turn, 25HC can enhance interferon production and inhibit uncontrolled inflammation. 25HC, being a functional antagonist of transcriptional SREBP, also inhibits IL-1 β and IL-1 formation by activated inflammasomes [27]. Such a dual role of 25HC in the immune response depends on the cellular and immunological context and can be explained by the need for an inflammatory response and protection from excessive inflammation when the production of anti-inflammatory agents leads to the restoration of homeostasis.

Interestingly, 25HC can prevent the penetration of viruses into the cell (HIV, Ebola virus, Zika virus, rabies virus, herpes simplex virus, SARS-CoV-2, etc.) [28–30]. This effect of 25HC is partly related to its ability to change the location of cholesterol by penetrating the membrane, thereby changing the properties of the membrane and lipid microdomains.

25-HYDROXYCHOLESTEROL AND THE CENTRAL NERVOUS SYSTEM

25-hydroxycholesterol is intensively synthesized by activated microglia, and its production can increase the formation of proinflammatory cytokines IL-1 β , IL-1 α , and tumor necrosis factor- α in response to the activation of TLR4 receptors by lipopolysaccharides [31]. In mice with *CH25H*-knockout microglia produces much less 25HC than microglia of wild-type animals [32].

25-hydroxycholesterol is a weak agonist of NMDA receptors of central synapses, which provide cognitive functions. Independently, 25HC only slightly enhances responses during activation of NMDA receptors; however, 25HC blocks the powerful potentiating effect of certain powerful positive allosteric modulators on NMDA receptors, particularly 24HC [33]. This effect of 25HC reduces the probability of excitotoxicity in hyperactivated NMDA receptors. The injection of lipopolysaccharide disrupts the long-term potentiation that is formed with the participation of NMDA receptors in the hippocampal regions of the mouse brain [32]. This impairment of long-term potentiation is not observed in mice with *CH25H*-knockout, which allows us to consider 25HC as a modulator of synaptic plasticity in neuroinflammation.

25-hydroxycholesterol is a ligand for LX receptors, which can form a complex with estrogen receptor- α [34]. 25HC can also directly activate estrogen receptors [16]. In turn, the activation of estrogen receptors by α -17 β -estradiol enhances long-term plasticity by increasing the clustering of AMPA receptors in the plasma membrane [35]. However, whether 25HC can act through estrogen receptors in the brain is unknown.

25-HYDROXYCHOLESTEROL AND NEUROMUSCULAR TRANSMISSION

macrophages play a key role in adaptive and innate immunity and are abundant in skeletal muscles, where they contact motor nerves and even neuromuscular junctions, promoting regeneration processes. 25HC is intensively produced by skeletal muscle macrophages in the presence of inflammation, which may indicate the functional significance of 25HC in the interaction of the immune system and skeletal muscles. Similar to the bidirectional concentration-dependent effect of 25HC on the immune system, the same effect was observed on the exocytosis of synaptic vesicles. High concentrations (1–10 μ mol) accelerate and low concentrations (0.01–0.10 μ mol) slow down this process [36].

25-hydroxycholesterol is a direct ligand for LX receptors. LX receptors are nuclear receptors; however, their expression has been detected in the plasma membrane of endothelial cells, thrombocytes [34, 37], and synaptic region of the axons of motor neurons [36]. 25HC in high concentration exerts its potentiating effect through the LX receptor-dependent pathway. Being in direct connection with estrogen receptor- α , LX receptors become active under the action of 25HC [36]. The same situation is observed when LX receptors are activated by an agonist in endothelial cells [34, 37]. LX receptors and estrogen receptor- α are located in lipid rafts, and the disruption of these microdomains abolished the potentiating effect of 25HC [36].

The deficiency in LX β receptors leads to the denervation and death of motor neurons [38], and the knockout of LX α and LX β receptors is characterized by the production of free radicals and lipid oxidation in some parts of the brain and sciatic nerves, which is accompanied by motor dysfunction [39]. Moreover, 25HC (1 μ mol) was found to increase the output of calcium ions from the endoplasmic reticulum through inositol triphosphate receptors, which is an additional contribution to the mobilization of synaptic vesicles to the active zone (exocytosis site) [36]. Zhong et al. discovered the opposite function of 25HC (1 μ mol): 25HC reduces the activity of the anti-apoptotic pathway ORP4L/ $G\alpha_{q/11}$ /PLC β 3/IP receptors/ Ca^{2+} in macrophages, which led to the induction of apoptosis [40]. Calcium-dependent protein kinases also enhance the mobilization of synaptic vesicles in Held synapses [41]. Similar data have been obtained regarding the neuromuscular synapse, where an increase in calcium output from intracellular depots under the action of 25HC (1 μ mol) leads to protein kinase activation with a subsequent increase in neurotransmitter release from synaptic vesicles during high-frequency activity [36].

Returning to estrogen receptor- α , their co-localization with the α -subunit and β -dimer of the G_i -protein determines the nongenomic effect of estradiol on endothelial cells

[42]. Interestingly, in neuromuscular synapses, high 25HC concentrations similarly influence the activity of estrogen receptor- α , leading to the dissociation of the $\beta\gamma$ -dimer from the G_i -protein, which potentiates the activity of phospholipase C [36].

An alternative realization of the 25HC effect on the neuromuscular synapse can be through ROS-dependent signaling. Calcium is released from the depot in response to 25HC and leads to an increase in reactive oxygen species (ROS) production by various mechanisms [36, 43]. Under the influence of 25HC (1 μmol), ROS production in the synaptic region and the concentration of hydrogen peroxide in the extracellular environment were increased. Interestingly, in this case, ROS play a signaling role because the level of lipid peroxidation under the action of 25HC was not affected [41]. In addition, like calcium, ROS can enhance the delivery of synaptic vesicles to exocytosis zones by activating protein kinase C [44–46].

25-HYDROXYCHOLESTEROL AND NEURODEGENERATIVE DISEASES

a high 25HC expression was observed in microglial cells in mice with an Alzheimer's disease. Interestingly, the microglia secreting the E4 variant of apolipoprotein E (which increases the risk of Alzheimer's disease) produce more 25HC and are more sensitive to the proinflammatory effect of 25HC than the microglia expressing apoE2 or apoE3 [31]. However, an increase in 25HC production by the microglia may also have a protective effect because 25HC reduces the clustering of the interferon- γ receptor in lipid rafts, thereby inhibiting the production of proinflammatory cytokines by microglia in response to interferon.

25-hydroxycholesterol is a potential marker of amyotrophic lateral sclerosis (ALS). ALS is characterized by progressive muscle atrophy, leading to fatality. In this pathology, 25HC concentration can reach micromolar concentrations in the spinal cord at the pre-onset disease stage in mice SOD1^{G93A} (model of ALS); however, this was not observed in patients with ALS, although CH25H overexpression was detected [47]. High concentrations, 5–30 μmol , can reduce survival and induce apoptosis of motor neurons [47, 48]. For example, 25HC activated mitochondrial-dependent apoptosis by stimulating GSK-3 β kinase. However, low concentrations, <1 μmol , have the opposite effects, increasing the survival of neurons [49].

Amyotrophic lateral sclerosis is characterized by a pronounced disturbance of lipid metabolism at the pre-onset stage [50, 51], including disturbances in membrane properties [52]. Lipid metabolism dysregulation is also observed in many other neurodegenerative diseases: spinal muscular atrophy [53], spinocerebellar ataxia [54], Huntington's disease [55], Parkinson's disease [56], and Alzheimer's disease [57, 58]. Recent studies have shown

increased content of glycosphingolipids and an imbalance between saturated and unsaturated fatty acids in ALS [59, 60]. In the model mice with mutation of a superoxide dismutase, high ceramide levels were found as a result of oxidative stress. In turn, ceramide is a lipid that induces programmed cell death. However, low ceramide concentrations have a neuroprotective effect and enhance axonal growth [61].

Impaired lipid metabolism, including sphingolipids and cholesterol, directly influences the state of lipid rafts, which determine the functioning of various proteins and protein complexes. Many studies have emphasized the important role of lipid rafts. For example, the level of caveolin-1 decreases in ALS, which leads to the impaired integrity of lipid rafts, contributing to disease progression [62]. In addition, a recent genomic study conducted by Zhang et al. [63] revealed changes in the content of 22 genes involved in protein expression associated with lipid rafts in patients with ALS, including ABCA1 (responsible for cholesterol efflux), CERS5 (involved in the formation of proapoptotic ceramide [64]), PLAA (phospholipase A activation), etc.

Synaptic hyperexcitability can potentiate the release of brain-derived neurotrophic factor and activation of TrkB receptors in ALS, followed by the induction of excitotoxicity and death of motor neurons [65]. In turn, TrkB receptors in combination with adenosine A2 receptors are localized in lipid rafts [66]. The extraction of cholesterol mainly from lipid rafts leads to a decrease in TrkB activity, thereby reducing excitotoxicity and death of motor neurons in ALS [65].

Changes in the properties of membranes in mice with a model of ALS at the pre-onset stage of the disease also support the important role of lipid rafts in ALS pathogenesis, indicating impaired integrity of lipid rafts. The use of various fluorescent labels disrupted the integrity of lipid rafts, increased the membrane fluidity, and destabilized the order of the lipid bilayer [52]. One of the possible causal factors of this phenomenon may be an increase in ceramide levels in the muscles, which leads to raft destabilization, for example, during hindlimb suspension [67–69]. In addition, a high ceramide level was observed in the membrane of mice with an ALS at the pre-onset stage [52].

Probably, lipid raft destabilization in ALS increases the level of extracellular choline concentration because of increased nonquantum secretion of acetylcholine [70]. In addition, the removal of cholesterol using the extracting agent methyl- β -cyclodextrin has the same effect; on the contrary, the addition of exogenous cholesterol reduces nonquantum secretion and choline levels [52]. An unregulated increase in acetylcholine level in ALS contributes to motor dysfunction and age-related morphological changes in the neuromuscular junction. For example, ALS also involved a disorder in the clustering of nicotinic acetylcholine receptors in the postsynaptic membrane [52, 71], which may explain the increase in amplitude and decrease in the rise time of miniature endplate potentials in mice with ALS [72].

Lipid raft destabilization can increase oxidative stress, for example, increased ROS production and increased lipid peroxidation in the neuromuscular junction are observed during the removal of cholesterol [73]. High hydroperoxide levels and lipid peroxidation were found in the homogenates of mouse muscles with the ALS model [50]. Interestingly, electrophilic aldehyde products that formed as a result of lipid peroxidation contribute to the aggregation of the SOD1 protein, and its mutation leads to ALS pathology [74].

As mentioned above, 25HC increases in ALS and affects the survival of motor neurons. In addition, 25HC induces lipid raft stabilization in neuromuscular synapses in mice with an ALS model at the pre-onset stage by reducing membrane fluidity and increasing its ordering. Moreover, 25HC prevents ceramide accumulation in the neuromuscular synapse. Interestingly, the synaptic membranes of mice with ALS have a significantly higher ability to bind 25HC than the membranes of wild-type mice [52]. Possibly, the number of binding sites for 25HC increases in ALS, including oxysterol-binding proteins, which can explain the high sensitivity of synaptic membranes of mice with ALS to 25HC. Data also show the involvement of oxysterol-binding proteins in ALS pathogenesis. For example, vesicle-associated protein B (VAPB) quantity, an important modulator of oxysterol-binding proteins, is reduced in the spinal cord in mice and patients with ALS at the pre-onset stage of the disease [75]. In addition, 25HC can regulate ceramide transport and cholesterol distribution in the membrane indirectly through oxysterol-binding proteins [76].

25-hydroxycholesterol can suppress synaptic changes in the neuromuscular synapse in ALS, that is, an increase in lipid peroxidation, extracellular choline levels, and disordered clustering of nicotinic acetylcholine receptors [52].

Based on the positive effect of 25HC on the properties of membranes, increased concentration in ALS can have a compensatory effect. Indeed, hypercholesterolemia has a protective effect on ALS, whereas substances that block cholesterol synthesis enhance this pathology [61, 77–80].

CONCLUSION

The class of cholesterol derivatives is diverse, and many of them have high biological activity. This study focused on 25-hydroxycholesterol, which is involved in the regulation of cellular cholesterol homeostasis and the functioning of the immune and nervous systems. Interestingly, 25-hydroxycholesterol has a concentration-dependent effect on both the immune and central nervous systems. Low 25-hydroxycholesterol concentrations promote the production of anti-inflammatory cells and improve synaptic plasticity. Higher 25-hydroxycholesterol concentrations can induce the secretion of proinflammatory cytokines and chemokines and inhibit synaptic activity, leading to synapse elimination.

25-hydroxycholesterol is produced by macrophages, including resident macrophages of the skeletal muscles. The 25-hydroxycholesterol level is increased in amyotrophic lateral sclerosis, which is characterized by the death of motor neurons and muscle atrophy. These data present the relationship between 25-hydroxycholesterol and neuromuscular transmission. Indeed, 25-hydroxycholesterol in micromolar concentrations enhances the recruitment of synaptic vesicles during the high-frequency activity of motor nerves. Membrane-associated LX receptors and estrogen receptor- α are involved in this effect of 25-hydroxycholesterol. These two receptors, forming a complex in lipid rafts, activate of the signaling pathway G_i -protein/ $\beta\gamma$ -dimer of G-protein/phospholipase C/ Ca^{2+} /protein kinase C. In addition, the Ca^{2+} -dependent increase in reactive oxygen species production under the action of 25-hydroxycholesterol also contributed to the enhancement of neuromuscular transmission.

Some studies have shown that high 25-hydroxycholesterol concentrations negatively affect amyotrophic lateral sclerosis; progression; however, a positive effect of 25-hydroxycholesterol on the properties of membranes at the pre-onset stage of the disease was also reported. Particularly, 25-hydroxycholesterol can restore synaptic anomalies such as increased membrane fluidity, ceramide accumulation, and decreased membrane ordering. In addition, 25-hydroxycholesterol reduces the high level of extracellular choline in amyotrophic lateral sclerosis, which can contribute to the fragmentation of neuromuscular synapses.

This review presents the functional role of 25-hydroxycholesterol in various body systems and allows us to consider cholesterol-25-hydroxylase as a therapeutic effect target in the future.

ADDITIONAL INFORMATION

Funding source. This work was supported by the Research Foundation Flanders (grant N 23-75-10022).

Competing interests. The authors declare that they have no competing interests.

Authors' contribution. G.F. Zakyranova — conducting experiments using optical and electrophysiological methods and data analysis, collecting and analyzing of literary sources, writing text and editing the article; A.N. Tsentssevitsky — conducting electrophysiological experiments and data analysis; E.A. Kuznetsova — conducting experiments using fluorescent labels and data analysis; S.M.F. Nghomsi — editing the article; A.R. Giniatullin — conducting electrophysiological experiments and data analysis; A.M. Petrov — designed of experimental plans and research management, collection and analysis of literary sources, writing of the text and editing of the article. All authors confirm that their authorship meets the international ICMJE criteria (all authors have made a significant contribution to the development of the concept,

research and preparation of the article, read and approved the final version before publication).

ДОПОЛНИТЕЛЬНО

Источник финансирования. Научное исследование проведено при поддержке Российского научного фонда (грант № 23-75-10022).

Конфликт интересов. Авторы декларируют отсутствие явных и потенциальных конфликтов интересов, связанных с публикацией настоящей статьи.

Вклад авторов. Г.Ф. Закирьянова — проведение экспериментов с использованием оптического и электрофизиологического методов и анализ данных, сбор и анализ литературных источников, написание текста

и редактирование статьи; А.Н. Ценцевицкий — проведение электрофизиологических экспериментов и анализ данных; Е.А. Кузнецова — проведение экспериментов с использованием флуоресцентных меток и анализ данных; С.М.Ф. Нгомси — редактирование статьи; А.Р. Гиниатуллин — проведение электрофизиологических экспериментов и анализ данных; А.М. Петров — разработка планов экспериментов и руководство исследованиями, сбор и анализ литературных источников, написание текста и редактирование статьи. Все авторы подтверждают соответствие своего авторства международным критериям ICMJE (все авторы внесли существенный вклад в разработку концепции, проведение исследования и подготовку статьи, прочли и одобрили финальную версию перед публикацией).

REFERENCES

- Lipowsky R, Sackmsnn E, editors. *Structure and dynamics of membranes*. 1st ed. Elsevier; 1995. 1052 p.
- Ma L, Nelson ER. Oxysterols and nuclear receptors. *Mol Cell Endocrinol*. 2019;484:42–51. doi: 10.1016/j.mce.2019.01.016
- Olivier E, Dutot M, Regazzetti A, et al. P2X7-pannexin-1 and amyloid β -induced oxysterol input in human retinal cell: role in age-related macular degeneration? *Biochimie*. 2016;127:70–78. doi: 10.1016/j.biochi.2016.04.014
- Bezine M, Namsi A, Sghaier R, et al. The effect of oxysterols on nerve impulses. *Biochimie*. 2018;153:46–51. doi: 10.1016/j.biochi.2018.04.013
- Gargiulo S, Gamba P, Testa G, et al. Molecular signaling involved in oxysterol-induced β_1 -integrin over-expression in human macrophages. *Int J Mol Sci*. 2012;13(11):14278–14293. doi: 10.3390/ijms131114278
- Yan D, Mäyränpää MI, Wong J, et al. OSBP-related protein 8 (ORP8) suppresses ABCA1 expression and cholesterol efflux from macrophages. *J Biol Chem*. 2008;283(1):332–340. doi: 10.1074/jbc.M705313200
- Kasimov MR, Fatkhrahmanova MR, Mukhutdinova KA, Petrov AM. 24S-hydroxycholesterol enhances synaptic vesicle cycling in the mouse neuromuscular junction: implication of glutamate NMDA receptors and nitric oxide. *Neuropharmacology*. 2017;117:61–73. doi: 10.1016/j.neuropharm.2017.01.030
- Mukhutdinova KA, Kasimov MR, Giniatullin AR, et al. 24S-hydroxycholesterol suppresses neuromuscular transmission in SOD1(G93A) mice: a possible role of NO and lipid rafts. *Mol Cell Neurosci*. 2018;88:308–318. doi: 10.1016/j.mcn.2018.03.006
- Mukhutdinova KA, Kasimov MR, Zakyrganova GF, et al. Oxysterol modulates neurotransmission via liver-X receptor/NO synthase-dependent pathway at the mouse neuromuscular junctions. *Neuropharmacology*. 2019;150:70–79. doi: 10.1016/j.neuropharm.2019.03.018
- Petrov AM, Pikuleva IA. Cholesterol 24-hydroxylation by CYP46A1: benefits of modulation for brain diseases. *Neurotherapeutics*. 2019;16(3):635–648. doi: 10.1007/s13311-019-00731-6
- Lütjohann D, Breuer O, Ahlborg G, et al. Cholesterol homeostasis in human brain: evidence for an age-dependent flux of 24S-hydroxycholesterol from the brain into the circulation. *Proc Natl Acad Sci U S A*. 1996;93(18):9799–9804. doi: 10.1073/pnas.93.18.9799
- Meaney S, Bodin K, Diczfalusy U, Björkhem I. On the rate of translocation in vitro and kinetics in vivo of the major oxysterols in human circulation: critical importance of the position of the oxygen function. *J Lipid Res*. 2002;43(12):2130–2135. doi: 10.1194/jlr.M200293-jlr200
- Lund EG, Xie C, Kotti T, et al. Knockout of the cholesterol 24-hydroxylase gene in mice reveals a brain-specific mechanism of cholesterol turnover. *J Biol Chem*. 2003;278(25):22980–22988. doi: 10.1074/jbc.M303415200
- Björkhem I, Lütjohann D, Diczfalusy U, et al. Cholesterol homeostasis in human brain: turnover of 24S-hydroxycholesterol and evidence for a cerebral origin of most of this oxysterol in the circulation. *J Lipid Res*. 1998;39(8):1594–1600.
- Blanc M, Hsieh WY, Robertson KA, et al. The transcription factor STAT-1 couples macrophage synthesis of 25-hydroxycholesterol to the interferon antiviral response. *Immunity*. 2013;38(1):106–118. doi: 10.1016/j.immuni.2012.11.004
- Liu Y, Wei Z, Ma X, et al. 25-hydroxycholesterol activates the expression of cholesterol 25-hydroxylase in an LXR-dependent mechanism. *J Lipid Res*. 2018;59(3):439–451. doi: 10.1194/jlr.M080440
- Bauman DR, Bitmansour AD, McDonald JG, et al. 25-hydroxycholesterol secreted by macrophages in response to Toll-like receptor activation suppresses immunoglobulin A production. *Proc Natl Acad Sci U S A*. 2009;106(39):16764–16769. doi: 10.1073/pnas.0909142106
- Karuna R, Christen I, Sailer AW, et al. Detection of dihydroxycholesterols in human plasma using HPLC-ESI-MS/MS. *Steroids*. 2015;99(Pt B):131–138. doi: 10.1016/j.steroids.2015.02.002
- Lehmann JM, Kliewer SA, Moore LB, et al. Activation of the nuclear receptor LXR by oxysterols defines a new hormone response pathway. *J Biol Chem*. 1997;272(6):3137–3140. doi: 10.1074/jbc.272.6.3137
- Zhu R, Ou Z, Ruan X, Gong J. Role of liver X receptors in cholesterol efflux and inflammatory signaling (review). *Mol Med Rep*. 2012;5(4):895–900. doi: 10.3892/mmr.2012.758
- Ouyang S, Mo Z, Sun S, et al. Emerging role of Insig-1 in lipid metabolism and lipid disorders. *Clin Chim Acta*. 2020;508:206–212. doi: 10.1016/j.cca.2020.05.042
- Radhakrishnan A, Ikeda Y, Kwon HJ, et al. Sterol-regulated transport of SREBPs from endoplasmic reticulum to Golgi: oxys-

- terols block transport by binding to INSIG. *Proc Natl Acad Sci U S A*. 2007;104(16):6511–6518. doi: 10.1073/pnas.0700899104
23. Sever N, Yang T, Brown MS, et al. Accelerated degradation of HMG CoA reductase mediated by binding of INSIG-1 to its sterol-sensing domain. *Mol Cell*. 2003;11(1):25–33. doi: 10.1016/s1097-2765(02)00822-5
24. Morens DM, Fauci AS. The 1918 influenza pandemic: insights for the 21st century. *J Infect Dis*. 2007;195(7):1018–1028. doi: 10.1086/511989
25. Kobasa D, Jones SM, Shinya K, et al. Aberrant innate immune response in lethal infection of macaques with the 1918 influenza virus. *Nature*. 2007;445(7125):319–323. doi: 10.1038/nature05495
26. Joseph SB, Bradley MN, Castrillo A, et al. LXR-dependent gene expression is important for macrophage survival and the innate immune response. *Cell*. 2004;119(2):299–309. doi: 10.1016/j.cell.2004.09.032
27. Reboldi A, Dang EV, McDonald JG, et al. Inflammation. 25-hydroxycholesterol suppresses interleukin-1-driven inflammation downstream of type I interferon. *Science*. 2014;345(6197):679–684. doi: 10.1126/science.1254790
28. Liu SY, Aliyari R, Chikere K, et al. Interferon-inducible cholesterol-25-hydroxylase broadly inhibits viral entry by production of 25-hydroxycholesterol. *Immunity*. 2013;38(1):92–105. doi: 10.1016/j.immuni.2012.11.005
29. Liu Y, Wei Z, Zhang Y, et al. Activation of liver X receptor plays a central role in antiviral actions of 25-hydroxycholesterol. *J Lipid Res*. 2018;59(12):2287–2296. doi: 10.1194/jlr.M084558
30. Yuan Y, Wang Z, Tian B, et al. Cholesterol 25-hydroxylase suppresses rabies virus infection by inhibiting viral entry. *Arch Virol*. 2019;164(12):2963–2974. doi: 10.1007/s00705-019-04415-6
31. Wong MY, Lewis M, Doherty JJ, et al. 25-hydroxycholesterol amplifies microglial IL-1 β production in an apoE isoform-dependent manner. *J Neuroinflammation*. 2020;17(1):192. doi: 10.1186/s12974-020-01869-3
32. Izumi Y, Cashikar AG, Krishnan K, et al. A proinflammatory stimulus disrupts hippocampal plasticity and learning via microglial activation and 25-hydroxycholesterol. *J Neurosci*. 2021;41(49):10054–10064. doi: 10.1523/JNEUROSCI.1502-21.2021
33. Linsenbardt AJ, Taylor A, Emnett CM, et al. Different oxysterols have opposing actions at N-methyl-D-aspartate receptors. *Neuropharmacology*. 2014;85:232–242. doi: 10.1016/j.neuropharm.2014.05.027
34. Ishikawa T, Yuhanna IS, Umetani J, et al. LXR β /estrogen receptor- α signaling in lipid rafts preserves endothelial integrity. *J Clin Invest*. 2013;123(8):3488–3497. doi: 10.1172/JCI66533
35. Clements L, Harvey J. Activation of oestrogen receptor α induces a novel form of LTP at hippocampal temporoammonic-CA1 synapses. *Br J Pharmacol*. 2020;177(3):642–655. doi: 10.1111/bph.14880
36. Zakyranova GF, Tsentsevitsky AN, Kuznetsova EA, Petrov AM. Immune-related oxysterol modulates neuromuscular transmission via non-genomic liver X receptor-dependent mechanism. *Free Radic Biol Med*. 2021;174:121–134. doi: 10.1016/j.freeradbiomed.2021.08.013
37. Unsworth AJ, Flora GD, Gibbins JM. Non-genomic effects of nuclear receptors: insights from the anucleate platelet. *Cardiovasc Res*. 2018;114(5):645–655. doi: 10.1093/cvr/cvy044
38. Bigini P, Steffensen KR, Ferrario A, et al. Neuropathologic and biochemical changes during disease progression in liver X receptor beta-/- mice, a model of adult neuron disease. *J Neuropathol Exp Neurol*. 2010;69(6):593–605. doi: 10.1097/NEN.0b013e3181df20e1
39. Hichor M, Sundaram VK, Eid SA, et al. Liver X receptor exerts a protective effect against the oxidative stress in the peripheral nerve. *Sci Rep*. 2018;8(1):2524. doi: 10.1038/s41598-018-20980-3
40. Zhong W, Pan G, Wang L, et al. ORP4L facilitates macrophage survival via G-protein-coupled signaling: ORP4L-/- mice display a reduction of atherosclerosis. *Circ Res*. 2016;119(12):1296–1312. doi: 10.1161/CIRCRESAHA.116.309603
41. Jin YH, Wu XS, Shi B, et al. Protein kinase C and calmodulin serve as calcium sensors for calcium-stimulated endocytosis at synapses. *J Neurosci*. 2019;39(48):9478–9490. doi: 10.1523/JNEUROSCI.0182-19.2019
42. Kumar P, Wu Q, Chambliss KL, et al. Direct interactions with G α i and G β \gamma mediate nongenomic signaling by estrogen receptor α . *Mol Endocrinol*. 2007;21(6):1370–1380. doi: 10.1210/me.2006-0360
43. Görlach A, Bertram K, Hudecova S, Krizanova O. Calcium and ROS: a mutual interplay. *Redox Biol*. 2015;6:260–271. doi: 10.1016/j.redox.2015.08.010
44. Giniatullin AR, Giniatullin RA. Dual action of hydrogen peroxide on synaptic transmission at the frog neuromuscular junction. *J Physiol*. 2003;552(Pt 1):283–293. doi: 10.1113/jphysiol.2003.050690
45. Cosentino-Gomes D, Rocco-Machado N, Meyer-Fernandes JR. Cell signaling through protein kinase C oxidation and activation. *Int J Mol Sci*. 2012;13(9):10697–10721. doi: 10.3390/ijms130910697
46. Ursan R, Odnoshivkina UG, Petrov AM. Membrane cholesterol oxidation downregulates atrial β -adrenergic responses in ROS-dependent manner. *Cell Signal*. 2020;67:109503. doi: 10.1016/j.cellsig.2019.109503
47. Dodge JC, Yu J, Sardi SP, Shihabuddin LS. Sterol auto-oxidation adversely affects human motor neuron viability and is a neuropathological feature of amyotrophic lateral sclerosis. *Sci Rep*. 2021;11(1):803. doi: 10.1038/s41598-020-80378-y
48. Kim SM, Noh MY, Kim H, et al. 25-hydroxycholesterol is involved in the pathogenesis of amyotrophic lateral sclerosis. *Oncotarget*. 2017;8(7):11855–11867. doi: 10.18632/oncotarget.14416
49. Choi YK, Kim YS, Choi IY, et al. 25-hydroxycholesterol induces mitochondria-dependent apoptosis via activation of glycogen synthase kinase-3 β in PC12 cells. *Free Radic Res*. 2008;42(6):544–553. doi: 10.1080/10715760802146062
50. Kim SM, Kim H, Kim JE, et al. Amyotrophic lateral sclerosis is associated with hypolipidemia at the presymptomatic stage in mice. *PLoS One*. 2011;6(3):e17985. doi: 10.1371/journal.pone.0017985
51. Chen X, Yazdani S, Piehl F, et al. Polygenic link between blood lipids and amyotrophic lateral sclerosis. *Neurobiol Aging*. 2018;67:202.e1–202.e6. doi: 10.1016/j.neurobiolaging.2018.03.022
52. Zakyranova GF, Giniatullin AR, Mukhutdinova KA, et al. Early differences in membrane properties at the neuromuscular junctions of ALS model mice: effects of 25-hydroxycholesterol. *Life Sci*. 2021;273:119300. doi: 10.1016/j.lfs.2021.119300
53. Deguise MO, Baranello G, Mastella C, et al. Abnormal fatty acid metabolism is a core component of spinal muscular atrophy. *Ann Clin Transl Neurol*. 2019;6(8):1519–1532. doi: 10.1002/acn3.50855
54. Darios F, Mochel F, Stevanin G. Lipids in the physiopathology of hereditary spastic paraplegias. *Front Neurosci*. 2020;14:74. doi: 10.3389/fnins.2020.00074
55. González-Guevara E, Cárdenas G, Pérez-Severiano F, Martínez-Lazcano JC. Dysregulated brain cholesterol metabolism is linked

- to neuroinflammation in huntington's disease. *Mov Disord*. 2020;35(7):1113–1127. doi: 10.1002/mds.28089
56. Fanning S, Selkoe D, Dettmer U. Parkinson's disease: proteinopathy or lipidopathy? *NPJ Parkinsons Dis*. 2020;6:3. doi: 10.1038/s41531-019-0103-7
57. Luchsinger JA, Cheng D, Tang MX, et al. Central obesity in the elderly is related to late-onset Alzheimer disease. *Alzheimer Dis Assoc Disord*. 2012;26(2):101–105. doi: 10.1097/WAD.0b013e318222f0d4
58. Tolppanen AM, Ngandu T, Kåreholt I, et al. Midlife and late-life body mass index and late-life dementia: results from a prospective population-based cohort. *J Alzheimers Dis*. 2014;38(1):201–209. doi: 10.3233/JAD-130698
59. Han X. Lipid alterations in the earliest clinically recognizable stage of Alzheimer's disease: implication of the role of lipids in the pathogenesis of Alzheimer's disease. *Curr Alzheimer Res*. 2005;2(1):65–77. doi: 10.2174/1567205052772786
60. Schengrund CL. Lipid rafts: keys to neurodegeneration. *Brain Res Bull*. 2010;82(1-2):7–17. doi: 10.1016/j.brainresbull.2010.02.013
61. Cutler RG, Pedersen WA, Camandola S, et al. Evidence that accumulation of ceramides and cholesterol esters mediates oxidative stress-induced death of motor neurons in amyotrophic lateral sclerosis. *Ann Neurol*. 2002;52(4):448–457. doi: 10.1002/ana.10312
62. Cooper-Knock J, Zhang S, Kenna KP, et al. Rare variant burden analysis within enhancers identifies CAV1 as an ALS risk gene. *Cell Rep*. 2020;33(9):108456. Corrected and republished from: *Cell Rep*. 2021. Vol. 34, N 5. doi: 10.1016/j.celrep.2020.108456
63. Zhang S, Cooper-Knock J, Weimer AK, et al. Genome-wide identification of the genetic basis of amyotrophic lateral sclerosis. *Neuron*. 2022;110(6):992–1008.e11. doi: 10.1016/j.neuron.2021.12.019
64. Levy M, Futerman AH. Mammalian ceramide synthases. *IUBMB Life*. 2010;62(5):347–356. doi: 10.1002/iub.319
65. Pradhan J, Noakes PG, Bellingham MC. The role of altered BDNF/TrkB signaling in amyotrophic lateral sclerosis. *Front Cell Neurosci*. 2019;13:368. doi: 10.3389/fncel.2019.00368
66. Mojsilovic-Petrovic J, Jeong GB, Crocker A, et al. Protecting motor neurons from toxic insult by antagonism of adenosine A2a and Trk receptors. *J Neurosci*. 2006;26(36):9250–9263. Corrected and republished from: *J Neurosci*. 2006;26(40):10079. doi: 10.1523/JNEUROSCI.1856-06.2006
67. Petrov AM, Kravtsova VV, Matchkov VV, et al. Membrane lipid rafts are disturbed in the response of rat skeletal muscle to short-term disuse. *Am J Physiol Cell Physiol*. 2017;312(5):C627–C637. doi: 10.1152/ajpcell.00365.2016
68. Petrov AM, Shalagina MN, Protopopov VA, et al. Changes in membrane ceramide pools in rat soleus muscle in response to short-term disuse. *Int J Mol Sci*. 2019;20(19):4860. doi: 10.3390/ijms20194860
69. Bryndina IG, Shalagina MN, Sekunov AV, et al. Clomipramine counteracts lipid raft disturbance due to short-term muscle disuse. *Neurosci Lett*. 2018;664:1–6. doi: 10.1016/j.neulet.2017.11.009
70. Petrov AM, Naumenko NV, Uzinskaya KV, et al. Increased non-quantal release of acetylcholine after inhibition of endocytosis by methyl- β -cyclodextrin: the role of vesicular acetylcholine transporter. *Neuroscience*. 2011;186:1–12. Corrected and republished from: *Neuroscience*. 2011;192:806. doi: 10.1016/j.neuroscience.2011.04.051
71. Sugita S, Fleming LL, Wood C, et al. VACHT overexpression increases acetylcholine at the synaptic cleft and accelerates aging of neuromuscular junctions. *Skelet Muscle*. 2016;6:31. doi: 10.1186/s13395-016-0105-7
72. Rocha MC, Pousinha PA, Correia AM, et al. Early changes of neuromuscular transmission in the SOD1(G93A) mice model of ALS start long before motor symptoms onset. *PLoS One*. 2013;8(9):e73846. doi: 10.1371/journal.pone.0073846
73. Petrov AM, Yakovleva AA, Zefirov AL. Role of membrane cholesterol in spontaneous exocytosis at frog neuromuscular synapses: reactive oxygen species-calcium interplay. *J Physiol*. 2014;592(22):4995–5009. doi: 10.1113/jphysiol.2014.279695
74. Esterbauer H, Schaur RJ, Zollner H. Chemistry and biochemistry of 4-hydroxynonenal, malonaldehyde and related aldehydes. *Free Radic Biol Med*. 1991;11(1):81–128. doi: 10.1016/0891-5849(91)90192-6
75. Teuling E, Ahmed S, Haasdijk E, et al. Motor neuron disease-associated mutant vesicle-associated membrane protein-associated protein (VAP) B recruits wild-type VAPs into endoplasmic reticulum-derived tubular aggregates. *J Neurosci*. 2007;27(36):9801–9815. doi: 10.1523/JNEUROSCI.2661-07.2007
76. Perry RJ, Ridgway ND. Oxysterol-binding protein and vesicle-associated membrane protein-associated protein are required for sterol-dependent activation of the ceramide transport protein. *Mol Biol Cell*. 2006;17(6):2604–2616. doi: 10.1091/mbc.e06-01-0060
77. Dupuis L, Corcia P, Fergani A, et al. Dyslipidemia is a protective factor in amyotrophic lateral sclerosis. *Neurology*. 2008;70(13):1004–1009. doi: 10.1212/01.wnl.0000285080.70324.27
78. Zheng Z, Sheng L, Shang H. Statins and amyotrophic lateral sclerosis: a systematic review and meta-analysis. *Amyotroph Lateral Scler Frontotemporal Degener*. 2013;14(4):241–245. doi: 10.3109/21678421.2012.732078
79. Krivoi II, Petrov AM. Cholesterol and the safety factor for neuromuscular transmission. *Int J Mol Sci*. 2019;20(5):1046. doi: 10.3390/ijms20051046
80. Ingre C, Chen L, Zhan Y, et al. Lipids, apolipoproteins, and prognosis of amyotrophic lateral sclerosis. *Neurology*. 2020;94(17):e1835–e1844. doi: 10.1212/WNL.0000000000009322

СПИСОК ЛИТЕРАТУРЫ

- Lipowsky R., Sackmann E., editors. Structure and dynamics of membranes. 1st ed. Elsevier, 1995. 1052 p.
- Ma L., Nelson E.R. Oxysterols and nuclear receptors // *Mol Cell Endocrinol*. 2019. Vol. 484. P. 42–51. doi: 10.1016/j.mce.2019.01.016
- Olivier E., Dutot M., Regazzetti A., et al. P2X7-pannexin-1 and amyloid β -induced oxysterol input in human retinal cell: Role in age-related macular degeneration? // *Biochimie*. 2016. Vol. 127. P. 70–78. doi: 10.1016/j.biochi.2016.04.014
- Bezine M., Namsi A., Sghaier R., et al. The effect of oxysterols on nerve impulses // *Biochimie*. 2018. Vol. 153. P. 46–51. doi: 10.1016/j.biochi.2018.04.013
- Gargiulo S., Gamba P., Testa G., et al. Molecular signaling involved in oxysterol-induced β_1 -integrin over-expression in human macrophages // *Int J Mol Sci*. 2012. Vol. 13, N 11. P. 14278–14293. doi: 10.3390/ijms131114278
- Yan D., Mäyränpää M.I., Wong J., et al. OSBP-related protein 8 (ORP8) suppresses ABCA1 expression and cholesterol efflux from

- macrophages // *J Biol Chem*. 2008. Vol. 283, N 1. P. 332–340. doi: 10.1074/jbc.M705313200
7. Kasimov M.R., Fatkhrahmanova M.R., Mukhutdinova K.A., Petrov A.M. 24S-hydroxycholesterol enhances synaptic vesicle cycling in the mouse neuromuscular junction: Implication of glutamate NMDA receptors and nitric oxide // *Neuropharmacology*. 2017. Vol. 117. P. 61–73. doi: 10.1016/j.neuropharm.2017.01.030
8. Mukhutdinova K.A., Kasimov M.R., Giniatullin A.R., et al. 24S-hydroxycholesterol suppresses neuromuscular transmission in SOD1(G93A) mice: a possible role of NO and lipid rafts // *Mol Cell Neurosci*. 2018. Vol. 88. P. 308–318. doi: 10.1016/j.mcn.2018.03.006
9. Mukhutdinova K.A., Kasimov M.R., Zakyrganova G.F., et al. Oxysterol modulates neurotransmission via liver-X receptor/NO synthase-dependent pathway at the mouse neuromuscular junctions // *Neuropharmacology*. 2019. Vol. 150. P. 70–79. doi: 10.1016/j.neuropharm.2019.03.018
10. Petrov A.M., Pikuleva I.A. Cholesterol 24-hydroxylation by CYP46A1: benefits of modulation for brain diseases // *Neurotherapeutics*. 2019. Vol. 16, N 3. P. 635–648. doi: 10.1007/s13311-019-00731-6
11. Lütjohann D., Breuer O., Ahlborg G., et al. Cholesterol homeostasis in human brain: evidence for an age-dependent flux of 24S-hydroxycholesterol from the brain into the circulation // *Proc Natl Acad Sci U S A*. 1996. Vol. 93, N 18. P. 9799–9804. doi: 10.1073/pnas.93.18.9799
12. Meaney S., Bodin K., Diczfalusy U., Björkhem I. On the rate of translocation in vitro and kinetics in vivo of the major oxysterols in human circulation: critical importance of the position of the oxygen function // *J Lipid Res*. 2002. Vol. 43, N 12. P. 2130–2135. doi: 10.1194/jlr.M200293-JLR200
13. Lund E.G., Xie C., Kotti T., et al. Knockout of the cholesterol 24-hydroxylase gene in mice reveals a brain-specific mechanism of cholesterol turnover // *J Biol Chem*. 2003. Vol. 278, N 25. P. 22980–22988. doi: 10.1074/jbc.M303415200
14. Björkhem I., Lütjohann D., Diczfalusy U., et al. Cholesterol homeostasis in human brain: turnover of 24S-hydroxycholesterol and evidence for a cerebral origin of most of this oxysterol in the circulation // *J Lipid Res*. 1998. Vol. 39, N 8. P. 1594–1600.
15. Blanc M., Hsieh W.Y., Robertson K.A., et al. The transcription factor STAT-1 couples macrophage synthesis of 25-hydroxycholesterol to the interferon antiviral response // *Immunity*. 2013. Vol. 38, N 1. P. 106–118. doi: 10.1016/j.immuni.2012.11.004
16. Liu Y., Wei Z., Ma X., et al. 25-Hydroxycholesterol activates the expression of cholesterol 25-hydroxylase in an LXR-dependent mechanism // *J Lipid Res*. 2018. Vol. 59, N 3. P. 439–451. doi: 10.1194/jlr.M080440
17. Bauman D.R., Bitmansour A.D., McDonald J.G., et al. 25-hydroxycholesterol secreted by macrophages in response to Toll-like receptor activation suppresses immunoglobulin A production // *Proc Natl Acad Sci U S A*. 2009. Vol. 106, N 39. P. 16764–16769. doi: 10.1073/pnas.0909142106
18. Karuna R., Christen I., Sailer A.W., et al. Detection of dihydroxycholesterols in human plasma using HPLC-ESI-MS/MS // *Steroids*. 2015. Vol. 99(Pt B). P. 131–138. doi: 10.1016/j.steroids.2015.02.002
19. Lehmann J.M., Kliewer S.A., Moore L.B., et al. Activation of the nuclear receptor LXR by oxysterols defines a new hormone response pathway // *J Biol Chem*. 1997. Vol. 272, N 6. P. 3137–3140. doi: 10.1074/jbc.272.6.3137
20. Zhu R., Ou Z., Ruan X., Gong J. Role of liver X receptors in cholesterol efflux and inflammatory signaling (review) // *Mol Med Rep*. 2012. Vol. 5, N 4. P. 895–900. doi: 10.3892/mmr.2012.758
21. Ouyang S., Mo Z., Sun S., et al. Emerging role of Insig-1 in lipid metabolism and lipid disorders // *Clin Chim Acta*. 2020. Vol. 508. P. 206–212. doi: 10.1016/j.cca.2020.05.042
22. Radhakrishnan A., Ikeda Y., Kwon H.J., et al. Sterol-regulated transport of SREBPs from endoplasmic reticulum to Golgi: oxysterols block transport by binding to INSIG // *Proc Natl Acad Sci U S A*. 2007. Vol. 104, N 16. P. 6511–6518. doi: 10.1073/pnas.0700899104
23. Sever N., Yang T., Brown M.S., et al. Accelerated degradation of HMG CoA reductase mediated by binding of INSIG-1 to its sterol-sensing domain // *Mol Cell*. 2003. Vol. 11, N 1. P. 25–33. doi: 10.1016/S1097-2765(02)00822-5
24. Morens D.M., Fauci A.S. The 1918 influenza pandemic: insights for the 21st century // *J Infect Dis*. 2007. Vol. 195, N 7. P. 1018–1028. doi: 10.1086/511989
25. Kobasa D., Jones S.M., Shinya K., et al. Aberrant innate immune response in lethal infection of macaques with the 1918 influenza virus // *Nature*. 2007. Vol. 445, N 7125. P. 319–323. doi: 10.1038/nature05495
26. Joseph S.B., Bradley M.N., Castrillo A., et al. LXR-dependent gene expression is important for macrophage survival and the innate immune response // *Cell*. 2004. Vol. 119, N 2. P. 299–309. doi: 10.1016/j.cell.2004.09.032
27. Reboldi A., Dang E.V., McDonald J.G., et al. Inflammation. 25-hydroxycholesterol suppresses interleukin-1-driven inflammation downstream of type I interferon // *Science*. 2014. Vol. 345, N 6197. P. 679–684. doi: 10.1126/science.1254790
28. Liu S.Y., Aliyari R., Chikere K., et al. Interferon-inducible cholesterol-25-hydroxylase broadly inhibits viral entry by production of 25-hydroxycholesterol // *Immunity*. 2013. Vol. 38, N 1. P. 92–105. doi: 10.1016/j.immuni.2012.11.005
29. Liu Y., Wei Z., Zhang Y., et al. Activation of liver X receptor plays a central role in antiviral actions of 25-hydroxycholesterol // *J Lipid Res*. 2018. Vol. 59, N 12. P. 2287–2296. doi: 10.1194/jlr.M084558
30. Yuan Y., Wang Z., Tian B., et al. Cholesterol 25-hydroxylase suppresses rabies virus infection by inhibiting viral entry // *Arch Virol*. 2019. Vol. 164, N 12. P. 2963–2974. doi: 10.1007/s00705-019-04415-6
31. Wong M.Y., Lewis M., Doherty J.J., et al. 25-hydroxycholesterol amplifies microglial IL-1 β production in an apoE isoform-dependent manner // *J Neuroinflammation*. 2020. Vol. 17, N 1. P. 192. doi: 10.1186/s12974-020-01869-3
32. Izumi Y., Cashikar A.G., K., et al. A proinflammatory stimulus disrupts hippocampal plasticity and learning via microglial activation and 25-hydroxycholesterol // *J Neurosci*. 2021. Vol. 41, N 49. P. 10054–10064. doi: 10.1523/JNEUROSCI.1502-21.2021
33. Linsenbardt A.J., Taylor A., Emmett C.M., et al. Different oxysterols have opposing actions at N-methyl-D-aspartate receptors // *Neuropharmacology*. 2014. Vol. 85. P. 232–242. doi: 10.1016/j.neuropharm.2014.05.027
34. Ishikawa T., Yuhanna I.S., Umetani J., et al. LXR β /estrogen receptor- α signaling in lipid rafts preserves endothelial integrity // *J Clin Invest*. 2013. Vol. 123, N 8. P. 3488–3497. doi: 10.1172/JCI66533
35. Clements L., Harvey J. Activation of oestrogen receptor α induces a novel form of LTP at hippocampal temporoammonic-CA1 synapses // *Br J Pharmacol*. 2020. Vol. 177, N 3. P. 642–655. doi: 10.1111/bph.14880
36. Zakyrganova G.F., Tsentssevitsky A.N., Kuznetsova E.A., Petrov A.M. Immune-related oxysterol modulates neuromuscular transmission via non-genomic liver X receptor-dependent mechanism // *Free Radic Biol Med*. 2021. Vol. 174. P. 121–134. doi: 10.1016/j.freeradbiomed.2021.08.013

- 37.** Unsworth A.J., Flora G.D., Gibbins J.M. Non-genomic effects of nuclear receptors: insights from the anucleate platelet // *Cardiovasc Res.* 2018. Vol. 114, N 5. P. 645–655. doi: 10.1093/cvr/cvy044
- 38.** Bigini P., Steffensen K.R., Ferrario A., et al. Neuropathologic and biochemical changes during disease progression in liver X receptor beta^{-/-} mice, a model of adult neuron disease // *J Neuropathol Exp Neurol.* 2010. Vol. 69, N 6. P. 593–605. doi: 10.1097/NEN.0b013e3181df20e1
- 39.** Hichor M., Sundaram V.K., Eid S.A., et al. Liver X receptor exerts a protective effect against the oxidative stress in the peripheral nerve // *Sci Rep.* 2018. Vol. 8, N 1. P. 2524. doi: 10.1038/s41598-018-20980-3
- 40.** Zhong W., Pan G., Wang L., et al. ORP4L facilitates macrophage survival via G-protein-coupled signaling: ORP4L^{-/-} mice display a reduction of atherosclerosis // *Circ Res.* 2016. Vol. 119, N 12. P. 1296–1312. doi: 10.1161/CIRCRESAHA.116.309603
- 41.** Jin Y.H., Wu X.S., Shi B., et al. Protein kinase c and calmodulin serve as calcium sensors for calcium-stimulated endocytosis at synapses // *J Neurosci.* 2019. Vol. 39, N 48. P. 9478–9490. doi: 10.1523/JNEUROSCI.0182-19.2019
- 42.** Kumar P., Wu Q., Chambliss K.L., et al. Direct interactions with G α i and G β γ mediate nongenomic signaling by estrogen receptor α // *Mol Endocrinol.* 2007. Vol. 21, N 6. P. 1370–1380. doi: 10.1210/me.2006-0360
- 43.** Görlach A., Bertram K., Hudecova S., Krizanova O. Calcium and ROS: a mutual interplay // *Redox Biol.* 2015. Vol. 6. P. 260–271. doi: 10.1016/j.redox.2015.08.010
- 44.** Giniatullin A.R., Giniatullin R.A. Dual action of hydrogen peroxide on synaptic transmission at the frog neuromuscular junction // *J Physiol.* 2003. Vol. 552(Pt 1). P. 283–293. doi: 10.1113/jphysiol.2003.050690
- 45.** Cosentino-Gomes D., Rocco-Machado N., Meyer-Fernandes J.R. Cell signaling through protein kinase C oxidation and activation // *Int J Mol Sci.* 2012. Vol. 13, N 9. P. 10697–10721. doi: 10.3390/ijms130910697
- 46.** Ursan R., Odnoshivkina U.G., Petrov A.M. Membrane cholesterol oxidation downregulates atrial β -adrenergic responses in ROS-dependent manner // *Cell Signal.* 2020. Vol. 67. P. 109503. doi: 10.1016/j.cellsig.2019.109503
- 47.** Dodge J.C., Yu J., Sardi S.P., Shihabuddin L.S. Sterol auto-oxidation adversely affects human motor neuron viability and is a neuropathological feature of amyotrophic lateral sclerosis // *Sci Rep.* 2021. Vol. 11, N 1. P. 803. doi: 10.1038/s41598-020-80378-y
- 48.** Kim S.M., Noh M.Y., Kim H., et al. 25-hydroxycholesterol is involved in the pathogenesis of amyotrophic lateral sclerosis // *Oncotarget.* 2017. Vol. 8, N 7. P. 11855–11867. doi: 10.18632/oncotarget.14416
- 49.** Choi Y.K., Kim Y.S., Choi I.Y., et al. 25-hydroxycholesterol induces mitochondria-dependent apoptosis via activation of glycogen synthase kinase-3 β in PC12 cells // *Free Radic Res.* 2008. Vol. 42, N 6. P. 544–553. doi: 10.1080/10715760802146062
- 50.** Kim S.M., Kim H., Kim J.E., et al. Amyotrophic lateral sclerosis is associated with hypolipidemia at the presymptomatic stage in mice // *PLoS One.* 2011. Vol. 6, N 3. P. e17985. doi: 10.1371/journal.pone.0017985
- 51.** Chen X., Yazdani S., Piehl F., et al. Polygenic link between blood lipids and amyotrophic lateral sclerosis // *Neurobiol Aging.* 2018. Vol. 67. P. 202. doi: 10.1016/j.neurobiolaging.2018.03.022
- 52.** Zakyryanova G.F., Giniatullin A.R., Mukhutdinova K.A., et al. Early differences in membrane properties at the neuromuscular junctions of ALS model mice: effects of 25-hydroxycholesterol // *Life Sci.* 2021. Vol. 273. P. 119300. doi: 10.1016/j.lfs.2021.119300
- 53.** Deguise M.O., Baranello G., Mastella C., et al. Abnormal fatty acid metabolism is a core component of spinal muscular atrophy // *Ann Clin Transl Neurol.* 2019. Vol. 6, N 8. P. 1519–1532. doi: 10.1002/acn3.50855
- 54.** Darios F., Mochel F., Stevanin G. Lipids in the physiopathology of hereditary spastic paraplegias // *Front Neurosci.* 2020. Vol. 14. P. 74. doi: 10.3389/fnins.2020.00074
- 55.** González-Guevara E., Cárdenas G., Pérez-Severiano F., Martínez-Lazcano J.C. Dysregulated brain cholesterol metabolism is linked to neuroinflammation in huntington's disease // *Mov Disord.* 2020. Vol. 35, N 7. P. 1113–1127. doi: 10.1002/mds.28089
- 56.** Fanning S., Selkoe D., Dettmer U. Parkinson's disease: proteinopathy or lipidopathy? // *NPJ Parkinsons Dis.* 2020. Vol. 6. P. 3. doi: 10.1038/s41531-019-0103-7
- 57.** Luchsinger J.A., Cheng D., Tang M.X., et al. Central obesity in the elderly is related to late-onset Alzheimer disease // *Alzheimer Dis Assoc Disord.* 2012. Vol. 26, N 2. P. 101–105. doi: 10.1097/WAD.0b013e318222f0d4
- 58.** Tolppanen A.M., Ngandu T., Kåreholt I., et al. Midlife and late-life body mass index and late-life dementia: results from a prospective population-based cohort // *J Alzheimers Dis.* 2014. Vol. 38, N 1. P. 201–209. doi: 10.3233/JAD-130698
- 59.** Han X. Lipid alterations in the earliest clinically recognizable stage of Alzheimer's disease: implication of the role of lipids in the pathogenesis of Alzheimer's disease // *Curr Alzheimer Res.* 2005. Vol. 2, N 1. P. 65–77. doi: 10.2174/1567205052772786
- 60.** Schengrund C.L. Lipid rafts: keys to neurodegeneration // *Brain Res Bull.* 2010. Vol. 82, N 1-2. P. 7–17. doi: 10.1016/j.brainresbull.2010.02.013
- 61.** Cutler R.G., Pedersen W.A., Camandola S., et al. Evidence that accumulation of ceramides and cholesterol esters mediates oxidative stress-induced death of motor neurons in amyotrophic lateral sclerosis // *Ann Neurol.* 2002. Vol. 52, N 4. P. 448–457. doi: 10.1002/ana.10312
- 62.** Cooper-Knock J., Zhang S., Kenna K.P., et al. Rare variant burden analysis within enhancers identifies CAV1 as an ALS risk gene // *Cell Rep.* 2020. Vol. 33, N 9. P. 108456. Corrected and republished from: *Cell Rep.* 2021. Vol. 34, N 5. P. 108730. doi: 10.1016/j.celrep.2020.108456
- 63.** Zhang S., Cooper-Knock J., Weimer A.K., et al. Genome-wide identification of the genetic basis of amyotrophic lateral sclerosis // *Neuron.* 2022. Vol. 110, N 6. P. 992–1008. doi: 10.1016/j.neuron.2021.12.019
- 64.** Levy M., Futerman A.H. Mammalian ceramide synthases // *IUBMB Life.* 2010. Vol. 62, N 5. P. 347–356. doi: 10.1002/iub.319
- 65.** Pradhan J., Noakes P.G., Bellingham M.C. The role of altered BDNF/TrkB signaling in amyotrophic lateral sclerosis // *Front Cell Neurosci.* 2019. Vol. 13. P. 368. doi: 10.3389/fncel.2019.00368
- 66.** Mojsilovic-Petrovic J., Jeong G.B., Crocker A., et al. Protecting motor neurons from toxic insult by antagonism of adenosine A2a and Trk receptors // *J Neurosci.* 2006. Vol. 26, N 36. P. 9250–9263. Corrected and republished from: *J Neurosci.* 2006. Vol. 26, N 40. P. 10079. doi: 10.1523/JNEUROSCI.1856-06.2006
- 67.** Petrov A.M., Kravtsova V.V., Matchkov V.V., et al. Membrane lipid rafts are disturbed in the response of rat skeletal muscle to short-term disuse // *Am J Physiol Cell Physiol.* 2017. Vol. 312, N 5. P. C627–C637. doi: 10.1152/ajpcell.00365.2016
- 68.** Petrov A.M., Shalagina M.N., Protopopov V.A., et al. Changes in membrane ceramide pools in rat soleus muscle in response to

- short-term disuse // *Int J Mol Sci.* 2019. Vol. 20, N 19. P. 4860. doi: 10.3390/ijms20194860
- 69.** Bryndina I.G., Shalagina M.N., Sekunov A.V., et al. Clomipramine counteracts lipid raft disturbance due to short-term muscle disuse // *Neurosci Lett.* 2018. Vol. 664. P. 1–6. doi: 10.1016/j.neulet.2017.11.009
- 70.** Petrov A.M., Naumenko N.V., Uzinskaya K.V., et al. Increased non-quantal release of acetylcholine after inhibition of endocytosis by methyl- β -cyclodextrin: the role of vesicular acetylcholine transporter // *Neuroscience.* 2011. Vol. 186. P. 1–12. Corrected and republished from: *Neuroscience.* 2011. Vol. 192. P. 806. doi: 10.1016/j.neuroscience.2011.04.051
- 71.** Sugita S., Fleming L.L., Wood C., et al. VACHT overexpression increases acetylcholine at the synaptic cleft and accelerates aging of neuromuscular junctions // *Skelet Muscle.* 2016. Vol. 6. P. 31. doi: 10.1186/s13395-016-0105-7
- 72.** Rocha M.C., Pousinha P.A., Correia A.M., et al. Early changes of neuromuscular transmission in the SOD1(G93A) mice model of ALS start long before motor symptoms onset // *PLoS One.* 2013. Vol. 8, N 9. P. e73846. doi: 10.1371/journal.pone.0073846
- 73.** Petrov A.M., Yakovleva A.A., Zefirov A.L. Role of membrane cholesterol in spontaneous exocytosis at frog neuromuscular synapses: reactive oxygen species-calcium interplay // *J Physiol.* 2014. Vol. 592, N 22. P. 4995–5009. doi: 10.1113/jphysiol.2014.279695
- 74.** Esterbauer H., Schaur R.J., Zollner H. Chemistry and biochemistry of 4-hydroxynonenal, malonaldehyde and related aldehydes // *Free Radic Biol Med.* 1991. Vol. 11, N 1. P. 81–128. doi: 10.1016/0891-5849(91)90192-6
- 75.** Teuling E., Ahmed S., Haasdijk E., et al. Motor neuron disease-associated mutant vesicle-associated membrane protein-associated protein (VAP) B recruits wild-type VAPs into endoplasmic reticulum-derived tubular aggregates // *J Neurosci.* 2007. Vol. 27, N 36. P. 9801–9815. doi: 10.1523/JNEUROSCI.2661-07.2007
- 76.** Perry R.J., Ridgway N.D. Oxysterol-binding protein and vesicle-associated membrane protein-associated protein are required for sterol-dependent activation of the ceramide transport protein // *Mol Biol Cell.* 2006. Vol. 17, N 6. P. 2604–2616. doi: 10.1091/mbc.e06-01-0060
- 77.** Dupuis L., Corcia P., Fergani A., et al. Dyslipidemia is a protective factor in amyotrophic lateral sclerosis // *Neurology.* 2008. Vol. 70, N 13. P. 1004–1009. doi: 10.1212/01.wnl.0000285080.70324.27
- 78.** Zheng Z., Sheng L., Shang H. Statins and amyotrophic lateral sclerosis: a systematic review and meta-analysis // *Amyotroph Lateral Scler Frontotemporal Degener.* 2013. Vol. 14, N 4. P. 241–245. doi: 10.3109/21678421.2012.732078
- 79.** Krivoi I.I., Petrov A.M. Cholesterol and the safety factor for neuromuscular transmission // *Int J Mol Sci.* 2019. Vol. 20, N 5. P. 1046. doi: 10.3390/ijms20051046
- 80.** Ingre C., Chen L., Zhan Y., et al. Lipids, apolipoproteins, and prognosis of amyotrophic lateral sclerosis // *Neurology.* 2020. Vol. 94, N 17. P. e1835–e1844. doi: 10.1212/WNL.0000000000009322

AUTHORS' INFO

* **Guzalia F. Zakyrganova**, Cand. Sci. (Biol.);
address: 2/31 Lobachevsky street, 420111 Kazan,
Russian Federation;
ORCID: 0000-0003-2949-0026;
eLibrary SPIN: 9856-1498;
e-mail: gffysiology@gmail.com

Andrei N. Tsentsevitsky, Cand. Sci. (Biol.);
ORCID: 0000-0002-4611-7509;
eLibrary SPIN: 2071-9047;
e-mail: atsen@list.ru

Arthur R. Giniatullin, Cand. Sci. (Biol.);
ORCID: 0000-0003-4789-1800;
eLibrary SPIN: 7614-5148;
e-mail: kvestor80@rambler.ru

Sonia M.F. Nghomsi;
ORCID: 0000-0001-8092-9431;
e-mail: danicastats@gmail.com

Eva A. Kuznetsova;
ORCID: 0000-0002-5581-7793;
e-mail: eva.korshak@mail.ru

Alexey M. Petrov, Cand. Sci. (Biol.);
ORCID: 0000-0002-1432-3455;
eLibrary SPIN: 7543-0918;
e-mail: apneurosci@gmail.com

* Corresponding author / Автор, ответственный за переписку

ОБ АВТОРАХ

* **Закирьянова Гузалия Фаритовна**, к.б.н.;
адрес: 420111, Российская Федерация, Казань,
ул. Лобачевского, д. 2/31;
ORCID: 0000-0003-2949-0026;
eLibrary SPIN: 9856-1498;
e-mail: gffysiology@gmail.com

Ценцевицкий Андрей Николаевич, к.б.н.;
ORCID: 0000-0002-4611-7509;
eLibrary SPIN: 2071-9047;
e-mail: atsen@list.ru

Гиниатуллин Артур Рауфович, к.б.н.;
ORCID: 0000-0003-4789-1800;
eLibrary SPIN: 7614-5148;
e-mail: kvestor80@rambler.ru

Нгомси Соня Маделен Фоген;
ORCID: 0000-0001-8092-9431;
e-mail: danicastats@gmail.com

Кузнецова Ева Андреевна;
ORCID: 0000-0002-5581-7793;
e-mail: eva.korshak@mail.ru

Петров Алексей Михайлович, д.б.н.;
ORCID: 0000-0002-1432-3455;
eLibrary SPIN: 7543-0918;
e-mail: apneurosci@gmail.com